# **Original Article**

# Effect of Streptococcus mutans on the flexural strength of resin-based restorative materials

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#### ABSTRACT

**Background:** There are a limited number of studies about the effects of microbial aging on the mechanical properties of restorative materials. Therefore, this study aimed to evaluate the effect of simulated aging with *Streptococcus mutans* on the flexural strength of different resin-based materials. **Materials and Methods:** This experimental study was performed on the blocks of different types of restorative materials including composite resin, giomer, and a resin-modified glass ionomer (RMGI). Moreover, three types of aging, such as 30-day storage in distilled water, *S. mutans*, and germ-free culture medium, were used in this study. The three-point bending flexural strength of the specimens before and after aging was measured according to the International Organization for Standardization-4049 standard. Data were analyzed by two-way ANOVA and *post hoc* Tukey's tests. A *P* < 0.05 was considered statistically significant.

Received: 23-Aug-2020 Revised: 27-Sep-2020 Accepted: 19-Apr-2021 Published: 21-Oct-2021

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E-mail: hvh\_haleh@yahoo. com **Results:** Results showed that the 30-day aging with the *S. mutans* significantly reduced the flexural strength of all three types of materials (P = 0.00). In all restorative materials, storage in a bacteria-free culture medium acted the same as distilled water, and there was no significant difference between these two solutions in terms of the flexural strength of the material, compared to the before-aging strength (P > 0.05). Furthermore, no significant difference was observed between *S. mutans*-based aging and distilled water aging regarding RMGI (P = 0.75).

**Conclusion:** It can be concluded that aging by *S. mutans* reduced the flexural strength in all three restorative materials.

Key Words: Aging, flexural strength, Streptococcus mutans

# INTRODUCTION

The complex nature of the oral environment provides several challenges to the restorative materials due to the presence of ions, enzymes, bacteria, pH, and temperature fluctuations. These factors can increase wear, surface roughness, color changes, and microleakage of restorations followed by a reduction in the mechanical properties.<sup>[1]</sup> The ability of restorative material to withstand these challenges

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Website: www.drj.ir www.drjjournal.net www.ncbi.nlm.nih.gov/pmc/journals/1480 is an important factor in their successful clinical performance. On the other hand, secondary caries and fractures are still the two main causes of restoration failure in the oral environment.<sup>[2-4]</sup> Despite the preventive measures, dental caries still occur and are among the most common chronic diseases<sup>[5]</sup> affecting a high percentage of the population.<sup>[6]</sup>

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**How to cite this article:** Haghi HV, Peeri-Dogaheh H, Fazlalizadeh S, Abazari M, Mohammadhosseini R. Effect of *Streptococcus mutans* on the flexural strength of resin-based restorative materials. Dent Res J 2021;18:90.

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Dental plaque is the major cause of dental caries, and Streptococcus mutans is one of the most important bacteria in cariogenic dental plaque. The virulence of these bacteria is mainly due to its high binding ability to biologic and restorative surfaces, as well as its acidogenic and aciduric nature.<sup>[7]</sup> The acid produced by these bacteria can not only demineralize the tooth structures but also is capable of destroying the resin-based restorative materials.<sup>[8]</sup> It has been shown that the tendency to form microbial plaque by these bacteria on the restoration surfaces is different among the restorative materials, and the resin-based materials show the greater tendency probably due to the release of un-polymerized resin monomers from these materials, which could accelerate the growth of these bacteria.<sup>[9]</sup>

Today, composite resins are widely used as restorative materials due to their esthetic and the ability of adhesion to tooth structure.<sup>[10,11]</sup> In addition to composite resins, other restorative materials that have resin components are also used to restore tooth defects. One of them is a group of resin-based materials called giomers. These light-cure materials have prereacted fillers, which have a similar composition to glass ionomer particles and are added to the resin matrix. These materials have simultaneously the advantages of composite resins' strength and esthetics, as well as glass ionomeric characteristics such as fluoride release and fluoride recharging.<sup>[12]</sup> Another group of dental materials having resins is resin-modified glass ionomers (RMGI), which are made by adding a small number of resin monomers to conventional glass ionomers. Therefore, they offer the improvement of optical properties, resistance to moisture, and initial better strength along with the benefits of adhesion to the tooth structure and the release of fluoride of conventional glass ionomers.<sup>[13]</sup>

Various studies have examined the effect of microbial plaque and *S. mutans* on the surface properties of restorative materials that showed some negative effects on the surface roughness and surface morphology of restorative materials;<sup>[7,9]</sup> however, few investigations have evaluated the effect of these microorganisms on the mechanical properties of restorative materials. Therefore, this study aimed to investigate the effect of microbial aging caused by *S. mutans* on the flexural strength of three groups of resin-containing restorative materials including composite resin, giomer, and RMGI. Moreover, it was attempted to compare their

effects with that of the water-based aging. The null hypothesis in this study was the lack of significant changes in the flexural strength of these three types of materials after 30 days of aging with *S. mutans*.

# **MATERIALS AND METHODS**

This experimental study was conducted on the resin based restorative material blocks. The restorative materials included nano-hybrid composite а resin (Charisma smart, Kulzer, Germany), а giomer (Beautifil II, Shofu Inc, Japan), and a RMGI (Fuji II LC, GC, Japan). According to the aging method, each material was divided into four subgroups of before aging, distilled water aging, S. mutans aging, and storage in a germ-free culture medium. It is worth mentioning that the flexural strength of the samples was measured in this study.

By conducting a pilot study, the mean flexural strength for giomer, resin composite, and RMGI was 105.26, 111.53, and 101.03 MPa, respectively. Considering the variance of 34.4, Type I error of 0.05 and 80% power, the sample size of 6 was selected based on the

formula of 
$$n = \frac{\gamma_{g,\alpha,l-\beta}}{\Lambda}, \Delta = \frac{\sum (\mu_l - \overline{\mu})^2}{\sigma^2}$$

A total of 72 composite blocks were fabricated. Table 1 summarizes the materials and their composition used in this study.

# **Specimen preparation**

Teflon molds with dimensions internal of 25 mm  $\times$  2 mm  $\times$  2 mm were utilized to prepare study specimens. The mold was placed on a celluloid strip over a glass-slab and fixed. It was then filled with restorative materials, the second piece of celluloid tape was placed on it, and the second slab was pressed to remove excess material. The slab was removed and the material was cured for 40 s in an overlapped manner along the entire length of the specimen using the LEDEX<sup>™</sup> WL-070 (Dentmate Technology, Taiwan) light-curing unit with the intensity of 1000 mW/cm<sup>2</sup>. Subsequently, the same exposure procedure was

Table 1:	Materials	used in	the	study	
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Material	Composition	Manufacturer
Charisma smart	Bis-GMA, barium aluminum fluoride glass, silicon dioxide	Kulzer GmbH, Hanau, Germany
Beautifil II	Bis-GMA, UDMA, Bis-MPEPP, TEGDMA, fluorosilicate glass	Shofu, Kyoto, Japan
Fuji II lc	HEMA, polyacrylic acid, water, fluoro-aluminum silicate glass	Gc, Tokyo, Japan

repeated on the other side of the specimen. A digital micrometer was employed to measure the dimensions of the prepared specimen (Digimatic, Mitutoyo, Japan), and the specimens with a difference of more than 0.01 mm were excluded from the study. To ensure the complete polymerization of the specimens, they were kept in distilled water over a period of 24-h incubation at 37°C.

#### Aging conditions

In total, four aging conditions were used in this study as follows:

# Before aging

In each group of restorative materials, the flexural strength of six blocks was measured right after 24-h of incubation.

#### Aging with distilled water

One group was kept in 200 ml of 37°C distilled water for 30 days.

# Aging with Streptococcus mutans

In this group, the specimens were immersed for 30 days in 200 ml of brain heart infusion (BHI) culture medium containing *S. mutans*.

#### Aging with brain heart infusion culture medium

To investigate the possible effects of BHI culture medium on materials' strength, a germ-free culture medium was used with a volume of 200 ml. The solutions were changed daily in all three aging groups.

# **Bacterial culture conditions**

S. mutans (ATCC700610, UA159) were provided from the research laboratory of the Department of Microbiology of Ardabil University of Medical Sciences, Ardabil, Iran, and were incubated aerobically at 37°C and 5%  $CO_2$  in the BHI medium (Sannyo, MCO-191C). The immersion solution containing 10<sup>7</sup> CFU/ml S. mutans was prepared daily.

### Measurement of flexural strength

The SANTAM STM-150 (Santam, Iran) universal testing machine was used to measure the flexural strength of specimens by a three-point bending test. To calculate the flexural strength, the following equation was used according to ISO-4049 standard:<sup>[14]</sup>

$$\sigma = \frac{3FL}{2bh^2}$$

Where F represents the maximum load in Newton, L signifies the distance between supports in millimeters, b indicates the specimen width in millimeters, and h presents the specimen thickness in millimeters.

#### **Statistical analysis**

Data were analyzed in SPSS software (version 23, statistical software IBM, USA). Due to the sample size of six in each group, the adjusted Kolmogorov–Smirnov test (Lilliefors test) was used to evaluate the data distribution. Moreover, to compare the flexural strength of materials under different aging conditions, two-way ANOVA and *post hoc* Tukey's test were utilized in this study. A P < 0.05 was considered statistically significant.

### RESULTS

The Lilliefors test results showed the normal distribution of the data in all groups (P > 0.05). Therefore, the two-way ANOVA test was used to compare the flexural strength values in different groups and showed significant differences between material groups (P < 0.001) and aging conditions (P < 0.001). However, there were no significant differences considering two factor of aging conditions and material types (P = 044). Table 2 summarizes the mean and standard deviations of the flexural strength values of the study groups. Furthermore, it demonstrates the results of the post hoc Tukey's test to compare the flexural strength values. The results showed that flexural strength values of all three types of materials significantly decreased after S. mutans aging condition (P = 0.001 for giomer, P = 0.003 for RMGI, P < 0.001 for composite resin). In addition, the values of flexural strength after S. mutans aging were lower than those in distilled water aging in the giomer (P < 0.001) and composite (P = 0.005)groups; however, they were the same in the RMGI group (P = 0.784).

#### DISCUSSION

In the present study, the effect of microbial aging caused by *S. mutans* was investigated on the flexural strength of resin-containing restorative materials including composite resin, giomer, and RMGI. According to the results, 30-day storage in conjunction with these bacteria significantly reduced the flexural strength of all three types of materials. In the experimental studies, aging by storage in different solutions, such as water,<sup>[15-17]</sup> is widely used for aging studies of restorative materials. However, this may not be sufficient to investigate the behavior of restorative materials in the oral environment, in which they are subjected to a variety of challenges.<sup>[18]</sup> One of the

Restorative materials	Aging conditions			
	Before aging	Aging with distilled water	Aging with Streptococcus mutans	Aging with BHI culture medium
Composite	147.26 (11.33) <sup>a, A</sup>	127.86 (11.48) <sup>a, A</sup>	89.69 (20.89) <sup>b, A</sup>	145.32 (11.02) <sup>a, A</sup>
Giomer	124.13 (8.78) <sup>a, A, C</sup>	105.71 (20.68) <sup>a, A</sup>	71.29 (23.88) <sup>b, A, C</sup>	119.50 (7.17) <sup>a, A</sup>
Glass ionomer	94.38 (10.33) <sup>a, B, C</sup>	71.07 (23.58) <sup>a, c, B</sup>	54.11 (8.79) <sup>b, c, B, C</sup>	80.56 (18.44) <sup>a, c, B</sup>

Table 2: Mean and standard devia	ions of flexural strength values
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Lowercase letters for each restorative material individually (rows) values denoted with same letters are not significantly different (P>0.05), Uppercase letters for each aging condition individually (column) values denoted with same letters are not significantly different (P>0.05). BHI: Brain heart infusion

challenges is microbial factors that should also be considered in aging studies. There are few studies in the literature regarding the effects of microbial aging on the strength of restorative materials. In 2018, Zhou et al. examined the effect of microbial aging on three common restorative materials including composite resin, giomer, and conventional glass ionomer. It should be noted that their results were consistent with the findings of the present study.[6] In 2019, a study was conducted by Algamaiah et al. on Bis-GMA-based composite material, ormocer, and an oxirane/acrylate experimental composite which yielded similar results for fracture toughness, except for oxirane/acrylate composite, the toughness of which did not change significantly after biofilm aging. This could be due to the hydrophobicity of the oxirane monomers.<sup>[19]</sup>

In our study, the resin components of the studied materials included monomers such as UDMA, Bis-GMA, TEGDMA, and HEMA. It has been shown that lactic acid and enzymes such as esterase produced by S. mutans can cause hydrolysis and chemical degradation of ester bonds in the resin matrix.<sup>[7,9]</sup> Therefore, reduction in flexural strength due to streptococcal aging in the composite and giomer groups can be attributed to resin degradation. Moreover, the resin/filler interface is affected by lactic acid. Destruction of interface leads to the separation of the mineral filler particles from the matrix that can further reduce the strength of the material.<sup>[6]</sup> In the case of the RMGI, which has a lower concentration of resin monomers than the other two groups, the resulted decrease in strength in addition to the reason for resin components' degradation can be due to an increase in mineral dissolution.

The lactic acid produced by *S. mutans* reduces the pH by 4.5,<sup>[20]</sup> and the mineral parts of the glass ionomer, such as  $AL_2$ ,  $O_3$ , and  $CaF_2$ , react with this acid.<sup>[21]</sup> It is well known that, in acidic environments, ionomeric materials are more susceptible to degradation than

resinous materials.<sup>[22]</sup> Therefore, despite the release of fluoride, which is expected to reduce microbial metabolism and plaque formation on the glass ionomeric and giomeric materials,<sup>[23]</sup> the flexural strength of RMGI decreased after microbial aging. This is probably due to the greater sensitivity of glass ionomer to dissolution and degradation in an acidic and hydrophilic environment in this study. Furthermore, some studies suggest that fluoride release of restorative materials is not a dominant or significant factor in plaque control; in addition, the released amount of fluoride is probably not sufficient due to washing out.<sup>[24]</sup>

In the present study, the effect of aging caused by *S. mutans* was compared with storage in distilled water. A germ-free culture medium was also used in a group to investigate the possible effects of BHI culture medium on flexural strength. In all types of the studied materials, storage in the BHI culture medium acted the same as distilled water, and there was no significant difference between these two conditions in terms of the flexural strength of the material. Therefore, the observed effects in *S. mutans* aging can be attributed to the direct effect of the bacteria.

After making a comparison between the strength of materials in the S. mutans and distilled water aging groups, the difference was significant, except for the RMGI material group. It is somewhat consistent with the results of a study performed by Zhou et al. In the aforementioned study, the composite resin showed no decrease in strength by retaining in water; however, after microbial aging, the strength decreased, which was in line with the results of our study. Nonetheless, in their study, glass ionomer and giomer showed significant reductions in strength by water aging. This difference in findings can be attributed to the use of RMGI in our study, which had lower solubility and higher moisture resistance, compared to conventional glass ionomer that used in the mentioned study.<sup>[6]</sup> Furthermore, the giomer used in the study carried out

by Zhou *et al.* was a flowable type; therefore, the decrease in strength after water aging can be explained by the lower filler rate. Regarding the effect of water-induced aging on the mechanical properties of composite resin materials, some studies have not shown a change in this regard,<sup>[6,25]</sup> whereas others reported a significant decrease. It seems that the change in strength is related to the duration of storage, and in 180 days of storage, all composite materials showed a reduced strength.<sup>[26,27]</sup> It is considered that the sensitivity to aqueous environments varies depending on the composition of the resin and its hydrophilicity.<sup>[28,29]</sup>

In the case of giomer and RMGI, the release of ions from fillers occurs in aqueous environments. As a result, the filler-matrix interface destroys due to the weakening of the filler surface that leads to a decrease in mechanical strength.<sup>[30]</sup> This strength reduction in the case of giomer is less than that in the RMGI since the acid-base reaction in the fillers of giomer takes place during the fabrication process. Therefore, the surface of the fillers readily has a modified layer that protects the central particles from the damaging effects of moisture.<sup>[30,31]</sup> In addition, the presence of a high percentage of fillers and different types of fillers in giomer is associated with more stable mechanical properties.<sup>[31]</sup>

It is noticeable that, in the RMGI group, although microbial aging can decrease the strength, compared to the initial strength before aging, it was not significantly different from the value of water aging. This finding probably indicates that the reduction in strength due to the *S. mutans* aging in the RMGI is somehow less than that in the other two materials. This finding can be attributed to the small effect of fluoride release.<sup>[23]</sup>

# CONCLUSION

Considering the limitations of the study, it can be concluded that aging caused by *S. mutans* in all three restorative materials reduced flexural strength. In the case of composite resin and giomer, the flexural strength after *S. mutans* aging was significantly different from water aging. Moreover, 30 days of storage in water showed no significant effect on the flexural strength of all three materials.

# Acknowledgment

This study was extracted from a research project No#2562 submitted to Ardabil University of Medical

Sciences, Ardabil, Iran, and financially supported by the Deputy of Research and Technology. The study protocol was approved by the Ethics Committee of Ardabil University of Medical Sciences, Ardabil, Iran (IR.ARUMS.REC.1398.444).

# Financial support and sponsorship Nil.

# **Conflicts of interest**

The authors of this manuscript declare that they have no conflicts of interest, real or perceived, financial or nonfinancial in this article.

# REFERENCES

- 1. Valinoti AC, Neves BG, da Silva EM, Maia LC. Surface degradation of composite resins by acidic medicines and pH-cycling. J Appl Oral Sci 2008;16:257-65.
- Mjör IA, Moorhead JE, Dahl JE. Reasons for replacement of restorations in permanent teeth in general dental practice. Int Dent J 2000;50:361-6.
- 3. Sarrett DC. Clinical challenges and the relevance of materials testing for posterior composite restorations. Dent Mater 2005;21:9-20.
- Ansari ZJ, Haghi HV. Secondary caries in the posterior teeth of patients presenting to the Department of Operative Dentistry, Shahid Beheshti Dental School. J Dent Sch Shahid Beheshti Med Sci Univ 2014;32:125-31.
- 5. Pitts NB, Zero DT, Marsh PD, Ekstrand K, Weintraub JA, Ramos-Gomez F, *et al.* Dental caries. Nat Rev Dis Primers 2017;3:17030.
- Zhou X, Wang S, Peng X, Hu Y, Ren B, Li M, *et al.* Effects of water and microbial-based aging on the performance of three dental restorative materials. J Mech Behav Biomed Mater 2018;80:42-50.
- Fúcio SB, Carvalho FG, Sobrinho LC, Sinhoreti MA, Puppin-Rontani RM. The influence of 30-day-old *Streptococcus mutans* biofilm on the surface of esthetic restorative materials – An *in vitro* study. J Dent 2008;36:833-9.
- Gonzalez-Bonet A, Kaufman G, Yang Y, Wong C, Jackson A, Huyang G, *et al.* Preparation of dental resins resistant to enzymatic and hydrolytic degradation in oral environments. Biomacromolecules 2015;16:3381-8.
- Beyth N, Bahir R, Matalon S, Domb AJ, Weiss EI. *Streptococcus mutans* biofilm changes surface-topography of resin composites. Dent Mater 2008;24:732-6.
- 10. Ansari ZJ, Moezzizadeh M, Haghi HV. Effect of different methods of blood decontamination on resin-resin micro-shear bond strength. Avicenna J Dent Res 2017;9:e60711.
- 11. Dietschi D, Shahidi C, Krejci I. Clinical performance of direct anterior composite restorations: A systematic literature review and critical appraisal. Int J Esthet Dent 2019;14:252-70.
- 12. Meena N. Giomer-The intelligent particle (new generation glass ionomer cement). Int J Dent Oral Health 2015;2:1-5.
- 13. Ellakuria J, Triana R, Mínguez N, Soler I, Ibaseta G, Maza J, *et al.* Effect of one-year water storage on the surface microhardness

of resin-modified versus conventional glass-ionomer cements. Dent Mater 2003;19:286-90.

- Yap AU, Eweis AH, Yahya NA. Dynamic and static flexural appraisal of resin-based composites: Comparison of the ISO and mini-flexural tests. Oper Dent 2018;43:E223-31.
- Okte Z, Villalta P, García-Godoy F, Lu H, Powers JM. Surface hardness of resin composites after staining and bleaching. Oper Dent 2006;31:623-8.
- Deepa CS, Krishnan VK. Effect of resin matrix ratio, storage medium, and time upon the physical properties of a radiopaque dental composite. J Biomater Appl 2000;14:296-315.
- Al Badr RM, Hassan HA. Effect of immersion in different media on the mechanical properties of dental composite resins. Int J Appl Dent Sci 2017;3:81-8.
- 18. Ferracane JL. Resin-based composite performance: Are there some things we can't predict? Dent Mater 2013;29:51-8.
- Algamaiah H, Danso R, Banas J, Armstrong SR, Whang K, Rawls HR, *et al.* The effect of aging methods on the fracture toughness and physical stability of an oxirane/acrylate, ormocer, and Bis-GMA-based resin composites. Clin Oral Investig 2020;24:369-75.
- Do D, Orrego S, Majd H, Ryou H, Mutluay MM, Xu HH, *et al.* Accelerated fatigue of dentin with exposure to lactic acid. Biomaterials 2013;34:8650-9.
- Wang L, Cefaly DF, Dos Santos JL, Dos Santos JR, Lauris JR, Mondelli RF, *et al. In vitro* interactions between lactic acid solution and art glass-ionomer cements. J Appl Oral Sci 2009;17:274-9.
- 22. Kantovitz KR, Pascon FM, Correr GM, Alonso RC, Rodrigues LK, Alves MC, *et al.* Influence of environmental conditions on

properties of ionomeric and resin sealant materials. J Appl Oral Sci 2009;17:294-300.

- Auschill TM, Arweiler NB, Brecx M, Reich E, Sculean A, Netuschil L. The effect of dental restorative materials on dental biofilm. Eur J Oral Sci 2002;110:48-53.
- Wiegand A, Buchalla W, Attin T. Review on fluoride-releasing restorative materials—fluoride release and uptake characteristics, antibacterial activity and influence on caries formation. Dent Mater 2007;23:343-62.
- 25. Sookhakiyan M, Tavana S, Azarnia Y, Bagheri R. Fracture toughness of nanohybrid and hybrid composites stored wet and dry up to 60 days. J Dent Biomater 2017;4:341-6.
- Bijelic-Donova J, Garoushi S, Lassila LV, Keulemans F, Vallittu PK. Mechanical and structural characterization of discontinuous fiber-reinforced dental resin composite. J Dent 2016;52:70-8.
- da Cunha LF, Saab RC, Costacurta AO, Baechtold MS, Zinelli RR, Fernandes AB, *et al.* Physicomechanical properties of different nanohybrid composites after aging: color stability, flexural strength, and microhardness. Braz J Oral Sci 2019;18:e191395.
- Mohammadi E, Pishevar L, Mirzakouchaki Boroujeni P. Effect of food simulating liquids on the flexural strength of a methacrylate and silorane-based composite. PLoS One 2017;12:e0188829.
- Lima A, Salvador M, Saraceni C. Influence of water storage periods on properties of resin materials. Dent Mater 2019;35:e23-4.
- Ilie N, Stawarczyk B. Evaluation of modern bioactive restoratives for bulk-fill placement. J Dent 2016;49:46-53.
- Garoushi S, Vallittu PK, Lassila L. Characterization of fluoride releasing restorative dental materials. Dent Mater J 2018;37:293-300.